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# INSTRUCTION MANUAL

## SI-SARC-CTS

*Optical Sarcomere Detection System for the SI-CTS*

Serial No. \_\_\_\_\_

[www.wpiinc.com](http://www.wpiinc.com)

121015



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## ABOUT THIS MANUAL

The following symbols are used in this guide:



This symbol indicates a CAUTION. Cautions warn against actions that can cause damage to equipment. Please read these carefully.



This symbol indicates a WARNING. Warnings alert you to actions that can cause personal injury or pose a physical threat. Please read these carefully.

NOTES and TIPS contain helpful information.



*Fig. 1 The camera is mounted on your inverted microscope and connected to your computer.*

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## INTRODUCTION

The **SI-H Optical Sarcomere Spacing System** includes a high speed microscope camera, the software necessary to measure and monitor the changing images and record data. It is designed for use with the **SI-CTS Cell Tester**, although a **Cell Tester** is not required for the **Optical Sarcomere Spacing System** to run. The microscope camera, with a maximum frame rate of 400 fps, mounts on your inverted microscope above the **Cell Tester** (if you use one). The video images of the cell are fed into the software for analysis. The software is based on the NIH open source, freeware "μManager."

A custom plugin allows the software to monitor sarcomere length. With the plugin, you may:

- Measure sarcomere spacing
- Run sarcomere calculations of 400Hz in the region of interest
- When used with a Cell Tester, record analog signal proportional to sarcomere length, and muscle parameters like force

## Parts List

After unpacking, verify that there is no visible damage to the sensor. Verify that all items are included:

- (1) USB3 Camera
- (1) 2μm Grating for calibration
- (1) Software Installation CD
- (1) Instruction Manual

## Unpacking

Upon receipt of this instrument, make a thorough inspection of the contents and check for possible damage. Missing cartons or obvious damage to cartons should be noted on the delivery receipt before signing. Concealed damage should be reported at once to the carrier and an inspection requested. Please read the section entitled "Claims and Returns" on page 23 of this manual. Please contact WPI Customer Service if any parts are missing at 941.371.1003 or customerservice@wpiinc.com.

**Returns:** Do not return any goods to WPI without obtaining prior approval (RMA # required) and instructions from WPI's Returns Department. Goods returned (unauthorized) by collect freight may be refused. If a return shipment is necessary, use the original container, if possible. If the original container is not available, use a suitable substitute that is rigid and of adequate size. Wrap the instrument in paper or plastic surrounded with at least 100mm (four inches) of shock absorbing material. For further details, please read the section entitled "Claims and Returns" on page 23 of this manual.

## SYSTEM SETUP

### Installing the Software

The WPI system uses the NIH open source  $\mu$ Manager program with a custom Sarcomere Detection plugin that is built for the USB3.0 camera. Before setting up the system, install the software.

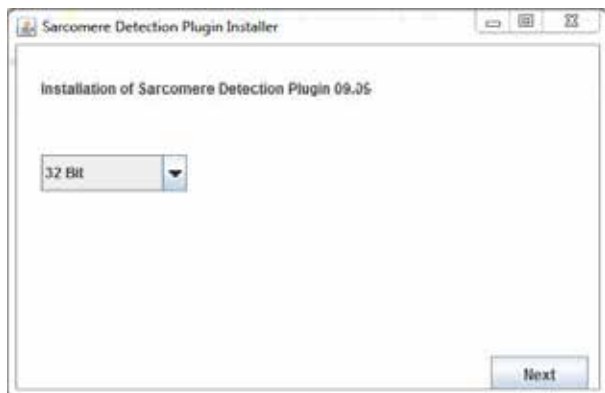
**NOTE:** Before you can successfully install the software, you must install the JAVA JDK for your operating system. Two files are included on your installation disk. Double click `jdk-8u5-windows-j586.exe` to install the 32-bit version or `jdk-8u5-windows-x64.exe` to install the 64-bit version.

1. Place the install CD into the CD drive on your computer. If the program does not launch automatically, use **Windows Explorer** to navigate to the CD drive. Double click the installation file **Installer.jar** (Fig. 2). The install program launches (Fig. 3), and all the necessary support programs will be installed, including:
  - WPI specific camera driver for 32-bit or 64-bit systems
  - $\mu$ Manager Environment support file
  - Sarcomere Detection plugin for  $\mu$ Manager.

Name	Date modified	Type
data	10/30/2014 1:42 PM	File folder
Installer	10/30/2014 11:38 ...	Executable Jar File

**Fig. 2** Double click on the Installer file to initiate the installation process.

2. Choose 32-Bit or 64-Bit from the drop down list, based on the operating system of your computer. Click **Next**. The Camera Installer appears (Fig. 4).



**Fig. 3** Choose 32-bit or 64-bit, based on your Operating System.

3. To install the camera software, click **Install**. The Camera Install Shield Wizard opens. Follow the instruction on screen. When the camera software installation

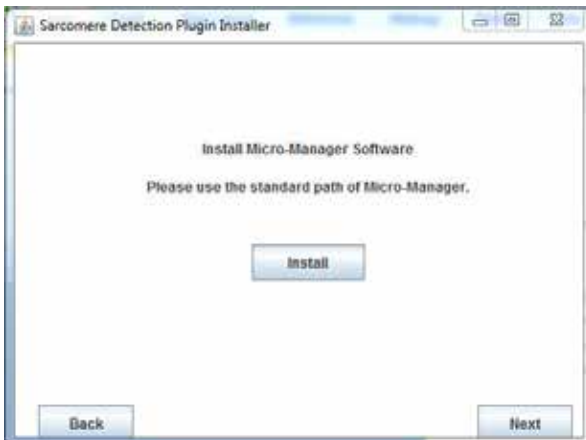
is finished, close the wizard. Click **Next** on the *Sarcomere Detection* plugin Installer window. The  $\mu$ Manager Software Installer appears (Fig. 5).



*Fig. 4 Click Install to open the camera software installation wizard.*

4. To install the  $\mu$ Manager software, click **Install** and follow the instructions on screen. For best results, do not change the default installation directory. When the installation is complete,  $\mu$ Manager opens. Close the  $\mu$ Manager program.

Then, click **Next** on the *Sarcomere Detection Plugin Installer* window. The *Sarcomere Detection Plugin installer* appears (Fig. 6).



*Fig. 5 Click Install to begin the installation of the  $\mu$ Manager Software and WPI driver for  $\mu$ Manager.*

5. To install of the *Sarcomere Detection* plugin, including the WPI camera driver for  $\mu$ Manager, choose the path for the location where the plugin will be installed. This



location should be the same place where  $\mu$ Manager is installed. Click **Install**. Then, confirm the installation path. When the **Sarcomere Detection** plugin files are installed in the designated location, a confirmation message appears.

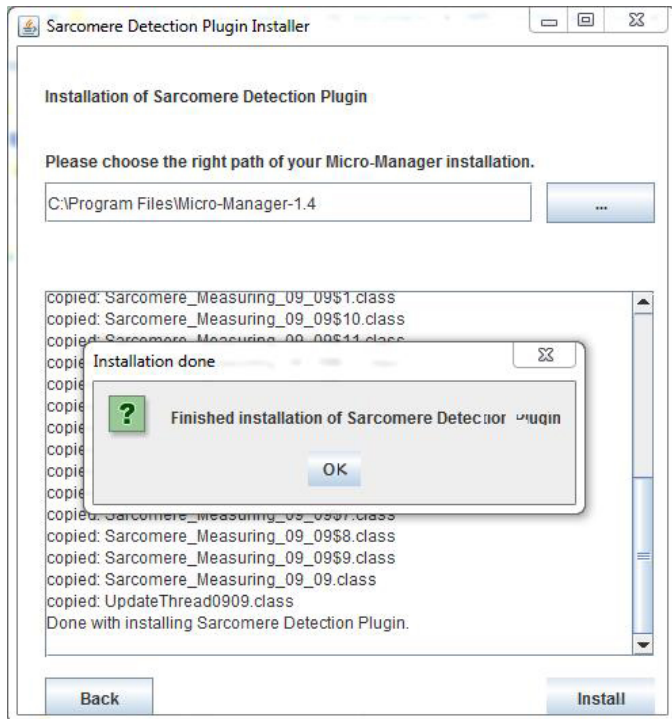


Fig. 6 When the installation of the Sarcomere Detection plugin is complete, a message appears.

6. Click **OK**.

## Configuring $\mu$ Manager for the USB 3.0 Camera

To use the camera in the  $\mu$ Manager environment, you must configure the camera.

1. To configure your camera, plug the camera into a USB 3.0 port on your computer. Then, open the  $\mu$ Manager program. The  **$\mu$ Manager Startup Configuration** window opens (Fig. 7).
2. If you have already configured a camera, you may select it from the drop down list. Otherwise, chose (none) to configure a new camera (Fig. 7).

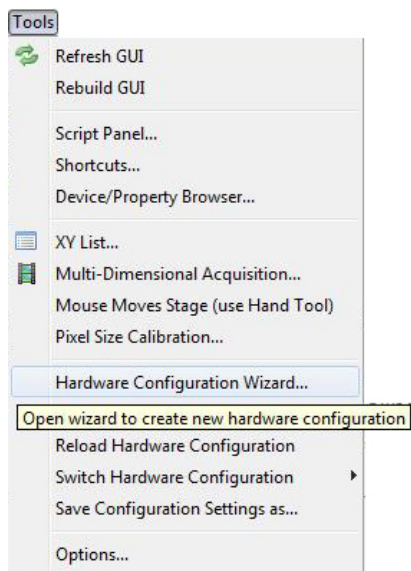


Fig. 7 (Left)  $\mu$ Manager Startup Configuration window

Fig. 8 (Right) Access the Hardware Configuration Wizard from the Tools menu.

3. On the  $\mu$ Manager main window menu bar, select the **Tools** menu and choose **Hardware Configuration Wizard** to open the camera configuration (Fig. 8). The Hardware Configuration Wizard opens.
4. Choose the **Create New Configuration** radio button and click **Next >** (Fig. 9). A list of available devices appears.

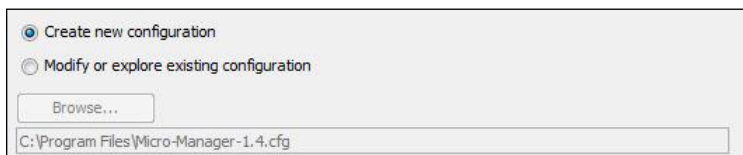


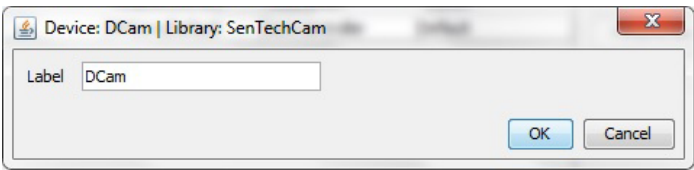
Fig. 9 First select the Create new configuration radio button.

5. To add the camera, search for the SenTechCam folder in the list of **Available Devices**. Open the folder and select **DCam | Sentech camera** (Fig. 10). Click **Add...**.



Fig. 10 Choose the SenTech camera from the list of available devices.

6. Give the device a name by entering a description into the Label field. Press



7. If the DCam was successfully added, it should be listed in the *Installed Devices* list in the upper left corner. Click **Next >** to finish the configuration. Complete the configuration by using the default values for steps 3–6.

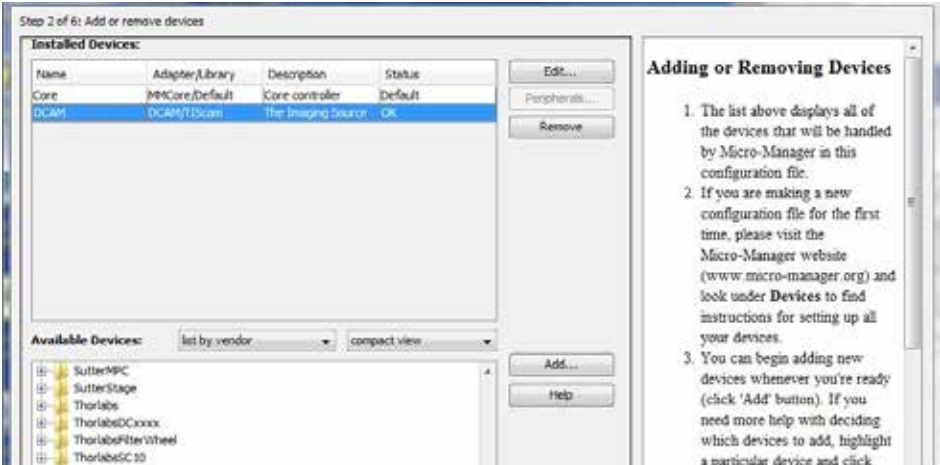


Fig. 11 The USB3 camera is listed in the Installed Devices section.

**NOTE:** For information on displaying and acquiring images, see the µManager instruction manual.

# Product Activation of Sarcomere Length Detection

WPI developed the custom plugin for use with the USB 3.0 camera to measure sarcomere length using  $\mu$ Manager. When you launch the *Sarcomere Detection* plugin for the first time, you are asked to activate the product to register your system.

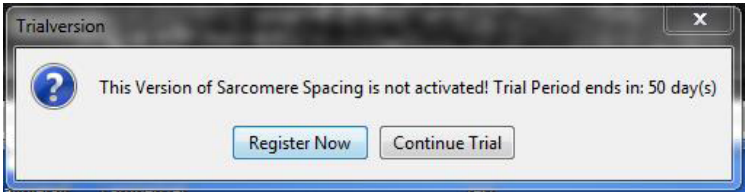


Fig. 12 Product activation is required

1. To register your product press **Register Now**. The *Product Registration and Activation* window opens (Fig. 13).

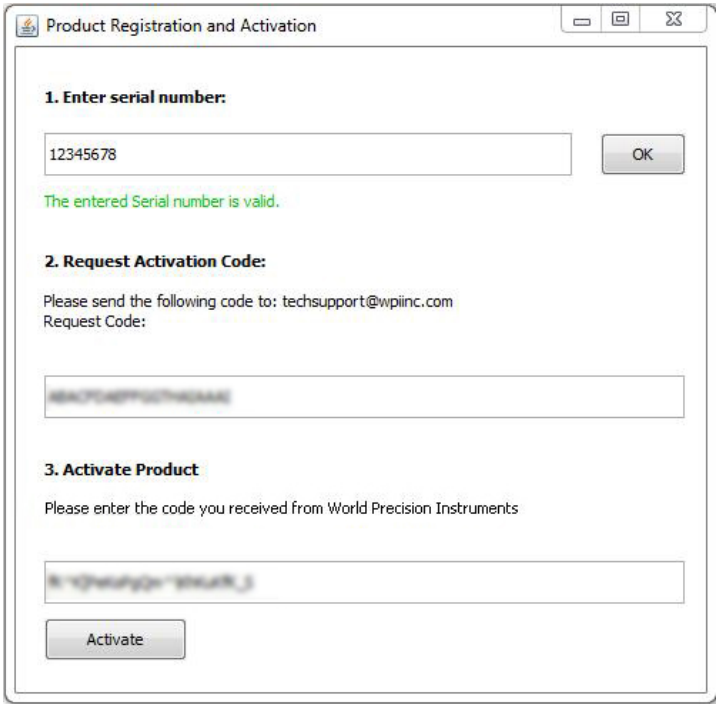
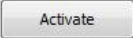


Fig. 13 The Product Registration and Activation window appears.

2. Enter your WPI serial number for the product. Press **OK** to validate the serial number. If the serial number is valid, a message appears under the serial

number field. A valid Serial number generates a **Request Activation Code**. Copy the code and send it to WPI at [techsupport@wpiinc.com](mailto:techsupport@wpiinc.com) to obtain a product key.

- When you receive your product key, enter it into the **Activate Product** field and press . A confirmation message appears indicating that your software was activated. Then, the **Sarcomere Detection** plugin is ready for use.

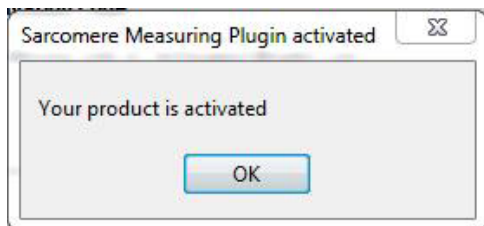


Fig. 14 The confirmation window indicates that the plugin is now activated.

## OPERATING INSTRUCTIONS

### Quick Start with $\mu$ Manager

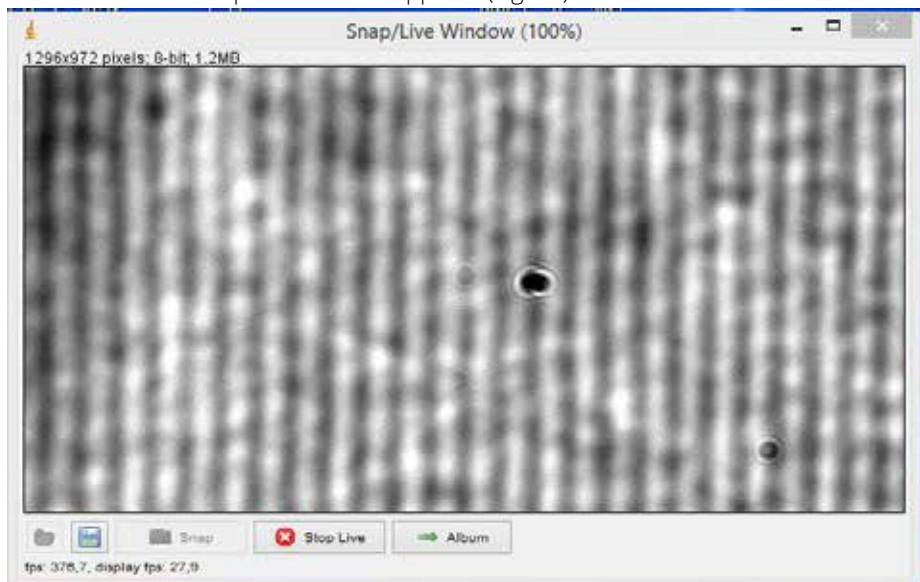
**NOTE:** Please refer to the  $\mu$ Manager Instruction Manual for complete direction on the use of  $\mu$ Manager. This section is included as a brief overview of using  $\mu$ Manager with your USB 3.0 camera.

- Change the **Exposure** time from 10ms to 0.1ms.





Fig. 15 Use these  $\mu$ Manager settings when using the USB 3.0 camera.

- Click on the  **Live** button to show your live image from the USB 3.0 camera. The Snap/Live window appears (Fig. 16).



*Fig. 16 The live viewing window of the USB 3.0 camera indicates the frame rate (fps). Here the ROI size of 1296x972 leads to 376.7 fps.*

- To increase the fps, change your ROI size in the live image. To do this, select  in the ImageJ window, and then select your desired ROI in the live window. Click on  to activate your selection. Watch as the frame rate changes depending on your ROI size.

**NOTE:** Imaging speeds greater than 1000 fps can be obtained depending on your ROI size.

## Using Sarcomere Detection Plugin with ImageJ

When  $\mu$ Manager opens, two main windows appears:

- $\mu$ Manager Main Window**—This is useful for running the camera
- ImageJ main window**—The Sarcomere Detection plugin for  $\mu$ Manager was installed for image processing. You may access the *Sarcomere Detection* plugin within ImageJ by selecting the *Plugins* menu and choosing *Sarcomere Detection* (Fig. 17). This brings up the most recently installed version of the *Sarcomere Detection* plugin.

**NOTE:** For information on using ImageJ, refer to the ImageJ manual.

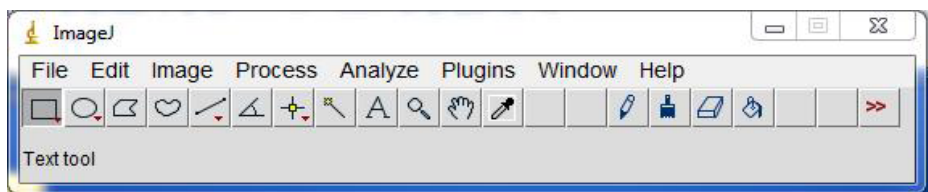


Fig. 17 You can access the Sarcomere Detection Plugin from the ImageJ main window.

1. To initiate sarcomere detection, open a sarcomere file (photo or film) or run  $\mu$ Manager for live detection of sarcomeres. Choose a region of interest (ROI) where you can easily distinguish the sarcomere striation (light/dark fields). A region of interest length of 10-15 sarcomeres is a reasonable field width. This can be seen in the Intensity/Sarcomere Length Chart when the **Sarcomere Detection** plugin is open (Fig. 22).

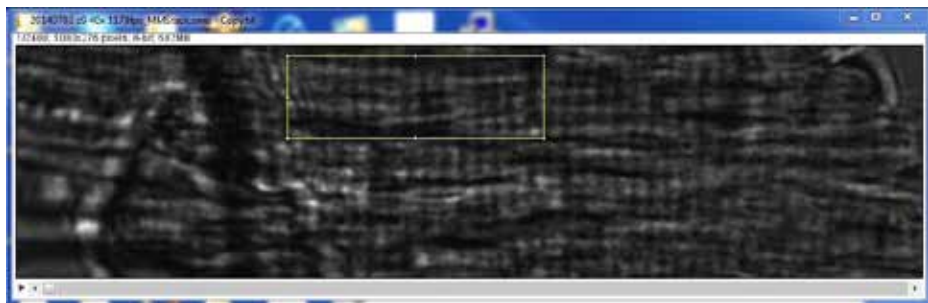


Fig. 18 The ImageJ display of a sarcomere picture was captured with  $\mu$ Manager. The yellow rectangle indicates the chosen region of interest. Image courtesy of Dr. Michael Kohlhaas, Universitäts Klinikum des Saarlandes, Homburg/Saar.

**TIP:** Always open your image in ImageJ and select your region of interest before opening the Sarcomere Detection software. Close the plugin before opening a different image.

2. To access the Sarcomere Detection software select the **Plugins** menu from the ImageJ main window menu bar, and choose **Sarcomere Detection**. The Sarcomere Detection software opens (Fig. 22).
3. Set up the parameters for the plugin and choose the filters you wish to use. See "Adjusting Plugin Parameters" on page 16.


**TIP:** The FFT Image J filter is more stable for contracting muscle cells.

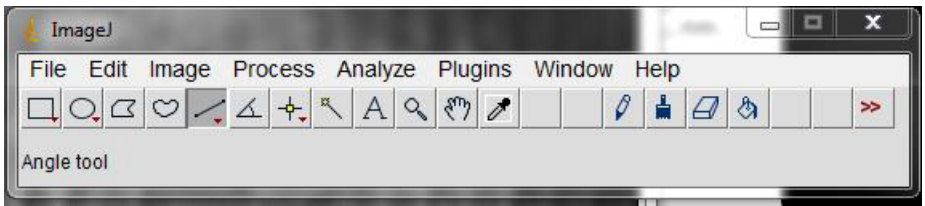
4. When you finish working with an image, close the plugin before you open another image.




## Calibrating the Objective

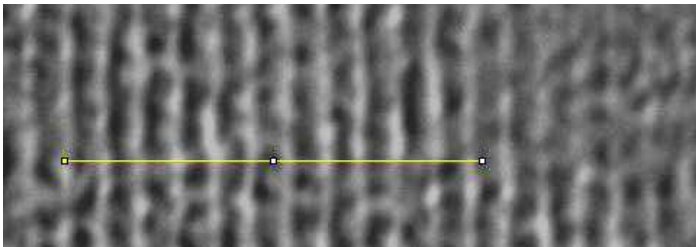
Before you begin making sarcomere measurements, you must calibrate your microscope eyepiece.

1. Place the  $2\mu\text{m}$  WPI grating (included) on your inverted microscope.
2. Open the image in  $\mu\text{Manager}$ . For the most accurate calibration measurement, zoom out to make the image as large as possible. Rotate the camera so that the grating lines are nearly vertical.
3. On ImageJ menu bar, select the straight line option from the angle tool (  ). Draw a horizontal line across your grating.



*Fig. 19 On the ImageJ window, the Straight Line option of the Angle Tool is selected.*

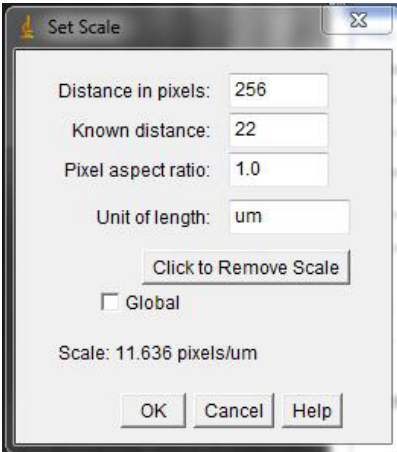
4. Count the grating lines that the ImageJ line crosses. The grating has a spacing of 500 grating lines/mm. So, the grating lines have a  $2\mu\text{m}$  spacing. In Fig. 20, we see 11 grating lines. For accurate calibration there should be at least 10 grating lines. For more information, press  on the **Set Scale** window (Fig. 21).



*Fig. 20 Draw a line in ImageJ across the field of view of the  $2\mu\text{m}$  grating that covers at least 10 grating lines.*

5. In ImageJ, select **Set Scale** from the **Analyze** menu to calculate the scaling factor. The **Set Scale** window appears. ImageJ automatically enters into the **Distance in pixels** field the distance measurement (in pixels) of the straight line that you drew.





*Fig. 21 The Set Scale window is used to calculate the scaling factor. In this example, the known distance is 22μm, and the scaling factor is 11.636 pixels/μm.*

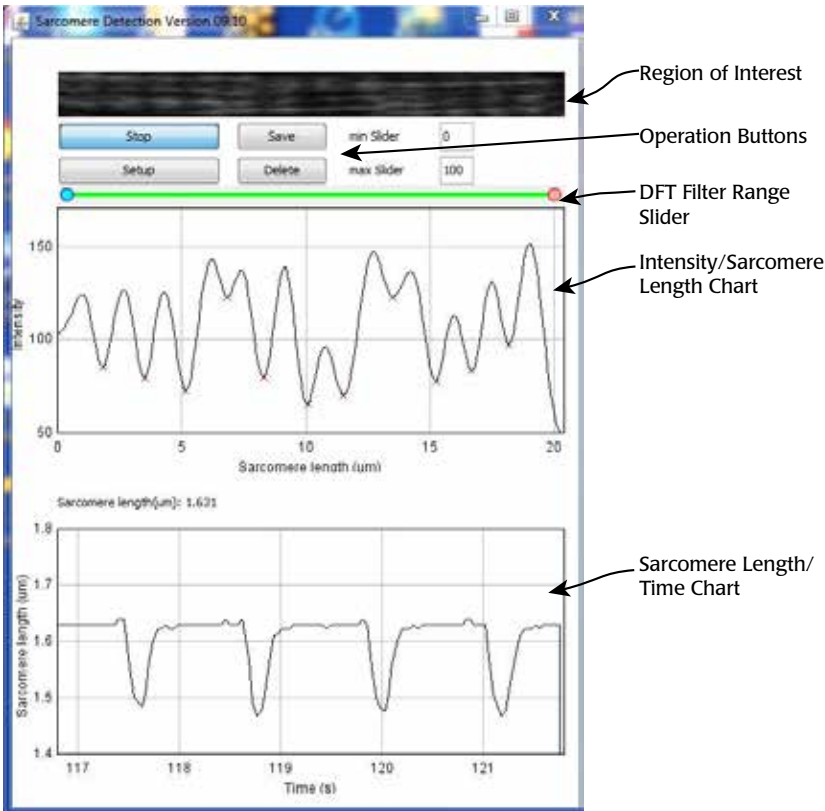
- Since the drawn line crosses 11 2μm grating lines, we know that the line we drew actually measures 22μm. Enter the actual length of the drawn line into the **Known Distance** field. The calculated scale factor is displayed at the bottom of the **Set Scale** window (11.636 pixels/μm).

**NOTE:** If you change the objective during sarcomere measurements, you will need to recalculate the scale factor, because the eyepiece calibration will change.

- When you are setting the Sarcomere Detection Plugin parameters, enter this scaling factor into the **Scale (pixels/μm)** field of the **Setup Sarcomere Detection** window (Fig. 25).

# Sarcomere Detection Window

This window shows the region of interest, an intensity chart and a sarcomere length chart. It also has four buttons for controlling the program.



*Fig. 22 The Sarcomere Detection plugin shows the region of interest, the intensity chart and a graph of the sarcomere length v. time.*

- Region of Interest**–The upper image is the display of the highlighted region of interest. This image is used to run the calculation of the sarcomere length.
- Start / Stop** –Use this button to start or stop the sarcomere length measuring. The button automatically toggles between Start and Stop.
- Setup** –Use this button to access the setup parameters for the plugin.
- Save** –Press this button to save the measured sarcomere length data. Pressing this button also resets the time buffer for saving data.
- Delete** – Press this button to clear the buffer so that new data may be acquired.

**NOTE:** Before you save data or delete the buffer, stop the data acquisition.

**DFT Filter Range Slider**—When the DFT filter is engaged, this slider is used to filter high and low frequencies. See “DFT” on page 20.

**Intensity/Sarcomere Length Chart**—The chart in the middle of the window displays the sinusoidal pattern of the chosen region of interest. Here the sarcomere length corresponds to the region of interest length. To improve the sinusoidal pattern (the light/dark cycle of the sarcomere striations), adjust the filter values using the setup parameters. See “Filter Settings” on page 18. The optimal sinusoidal pattern is achieved when you see one red X for each downward inflection of the sinusoidal pattern.

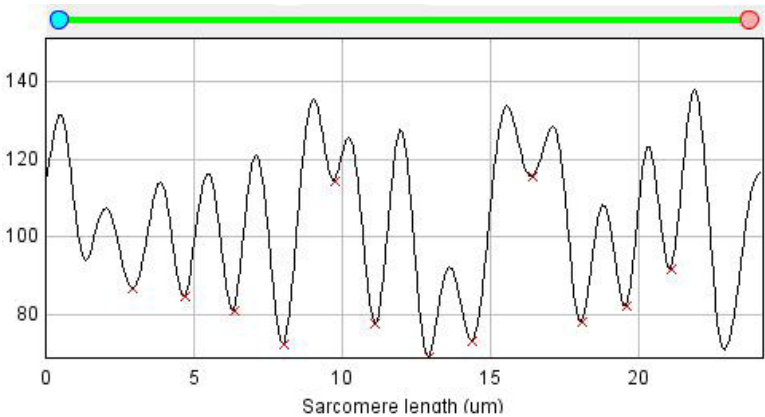


Fig. 23 The Intensity/Sarcomere Length Chart shows the sinusoidal pattern of the region of interest.

**Sarcomere Length/Time Chart**—The bottom chart displays the measured sarcomere length change over the measured time (Fig. 24).

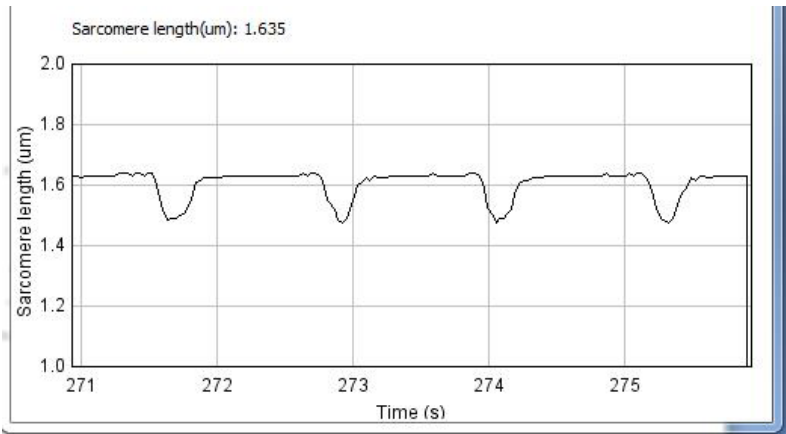
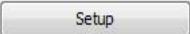


Fig. 24 Sarcomere length change during a 1 Hz contraction of a cardiac myocyte.

# Adjusting Plugin Parameters

Before you begin making measurements, you may adjust the plugin parameters. To do this you press . The *Setup Sarcomere Detection* window appears.

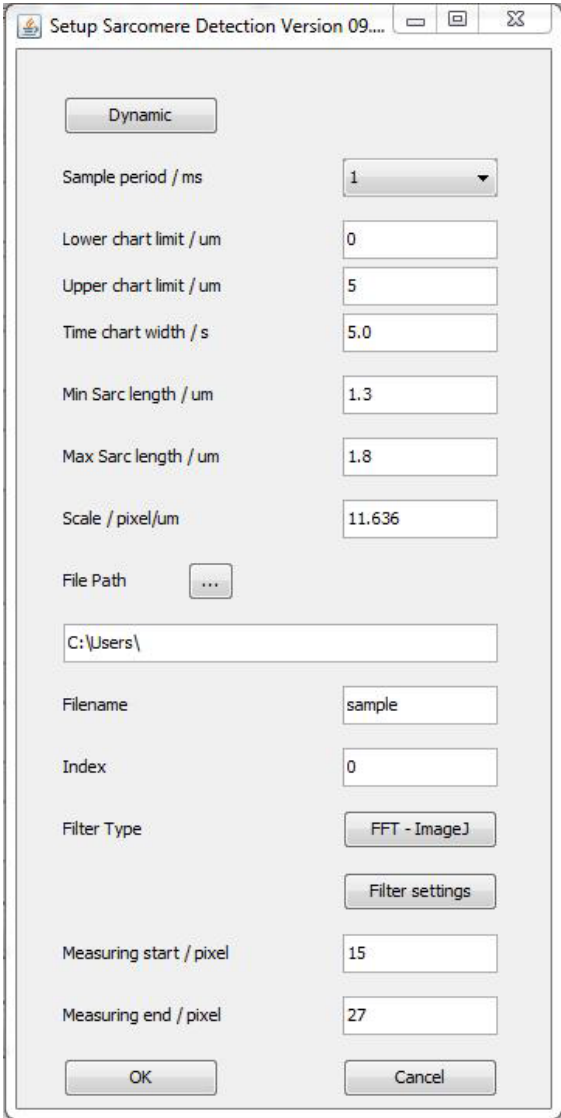
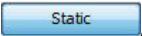
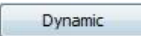


Fig. 25 Use the Setup Sarcomere Detection window to define the plugin parameters.

  –Press this button to toggle between static mode and dynamic mode. It may be useful to toggle between the modes when you switch from running tests on resting muscle condition (static mode) and contracting muscle condition (dynamic mode). If you are using a large region of interest with imperfections in your image, you will see better results with static mode. You can even use the Min and Max Sarc Length parameters to filter out these image imperfections.

**Sample Period (ms)**–Select your sample period (in milliseconds) from the drop down list. This defines the waiting period between two measurements.

**NOTE:** A measurement will not begin until a previous measurement has finished. This could happen if the measurement takes longer than the sample period, which could happen if the region of interest is too large.

**Lower Chart Limit ( $\mu\text{m}$ )**–Set the minimum value in microns of the Y-axis for the *Sarcomere Length/Time Chart*.

**Upper Chart Limit ( $\mu\text{m}$ )**–Set the maximum value in microns for the Y-axis for the *Sarcomere Length/Time Chart*.


**Time Chart Width (s)**–Set the total width in seconds of the X-axis for *Sarcomere Length/Time Chart*.

**Min Sarc Length ( $\mu\text{m}$ )** – Set the minimum sarcomere boundary to be detected. A sarcomere length smaller than this threshold is ignored.

**Max Sarc Length ( $\mu\text{m}$ )** – Set the maximum sarcomere boundary to be detected. A sarcomere length larger than this threshold is ignored.

**TIP:** When sarcomere length is outside the range set by the minimum and maximum thresholds, a question mark displays for the sarcomere length on the Sarcomere Detection Window. To see a value, expand your detection range (Min/Max Sarc Length settings) or select a larger region of interest.

**Scale (pixel/ $\mu\text{m}$ )**–Set the calibrating factor to compare pixels with microns. See “Calibrating the Objective” on page 12.

**File Path**  –Press this button to open the folder browser and select the location of your saved files. After you select the folder location, the file path appears in the text field. If you need to create a new folder, use Windows Explorer.

**File Name**–Enter the name for your new data file where the measurement data will be stored. If you do not change the file name here, the existing name will be appended with a number that increments automatically each time measurements are saved.

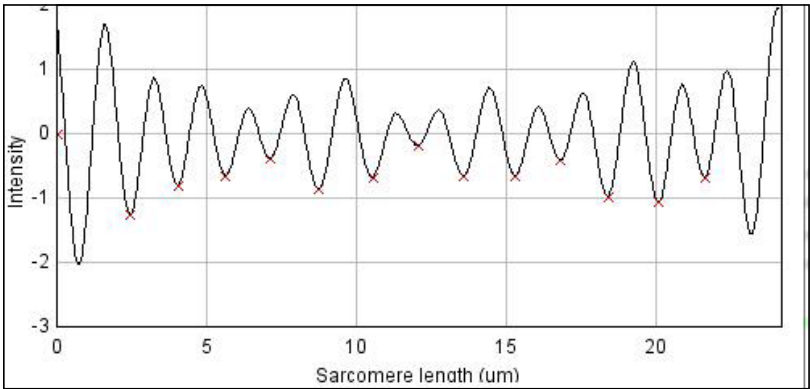
**Index**–This is the starting number used when the file name is automatically incremented. In the example image, the first file name will be *sample0*.

**Filter Type**–Filters are used for image processing, and each type has its own settings which are detailed later. Click on the **Filter Type** button to toggle between the filter option. The filter type options include:

- **FFT-ImageJ**–This is an ImageJ based FFT image filter (page 18).
- **DFT** is a Discrete Fourier Transform, which filters out a selected frequency range (page 20).

**Measuring Start (pixel)**–Set a length in pixel for the zero point. The program skips these pixels on the left side of the region of interest and does not make measurements on them.

**Measuring End (pixel)**–Set a length in pixels for the end point. This is an inset from the right side of the region of interest. Pixels within the designated border region are not used for measurements.



*Fig. 26 This chart shows that the start point is not at the zero point (left) of the chart's X-axis, and the end point is not at the far right of the chart X-axis. Each red X indicates that a measurement was made. In this example, measurements start around 2.5µm and end around 22.5µm.*

## Filter Settings

**NOTE:** FFT ImageJ filter settings need to be activated when you are running the sarcomere detection.

### FFT-ImageJ

This filter is used to sharpen an image by removing blur caused by things like movement artifacts. Two options for this filter, which are often used together, allow you to filter out the largest structures and/or filter out the smallest structures. When you select the *FFT-ImageJ* filter type, the **Filter settings** button appears. Click the button to open the *FFT Bandpass Filter* window (Fig. 28) and set your parameters for the filter. After you set your parameters, click **OK**. For more information on this filter type, press **Help**.



*Fig. 27 When the FFT ImageJ filter type is selected, the Filter Setting button appears.*

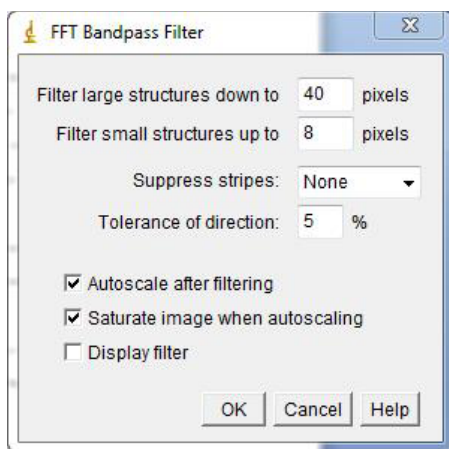


Fig. 28 Use this window to establish the parameters for your filter.

**Filter Large Structures Down to**—Enter a number of pixels here. Structures comprised of more pixels than this threshold are filtered. This filter corresponds with the suppression of low frequencies in the image. It is typically chosen to filter out smooth variations like movement artifacts in the background. Values greater than the chosen filter value are suppressed. To suppress movement artifacts, increase this filter value.

**Filter Small Structures Up to**—Enter a number of pixels here. Structures comprised of fewer pixels than this threshold are filtered. This filter corresponds with the suppression of high frequencies in the image and determines the amount of smoothing. Objects in the image smaller than this size are strongly attenuated. To remove movement artifacts increase the filter value. Resting sharp peak values in the image are also filtered out by increasing this value.

**NOTE:** Increasing the region of interest size/width may improve the detection quality.

**Suppress Stripes**—Use this drop down list to choose whether or not to eliminate horizontal or vertical stripes.

**Tolerance of Direction**—If you use *Suppress Stripes*, you may set a tolerance value. Higher values remove shorter stripes and stripes that are running under an angle with respect to the chosen direction (horizontal or vertical).

**Autoscale after Filtering**—By selecting this checkbox, you turn on autoscaling. The lowest intensity is set to 0, and the highest intensity is set to 255.

**Saturate image when Autoscaling** allows some intensities to go into saturation and produces a better visual contrast. Saturate will only produce an effect when autoscaling is enabled.


**Display Filter** shows the filter that is generated.

## DFT

The DFT (Discrete Fourier Transform) filter allows you to filter low and high frequencies in the image during the sarcomere length measurement. To activate this filter, select DFT as the filter type in the **Setup Sarcomere Detection** window. When you close the **Setup Sarcomere Detection** window, the filter is active.

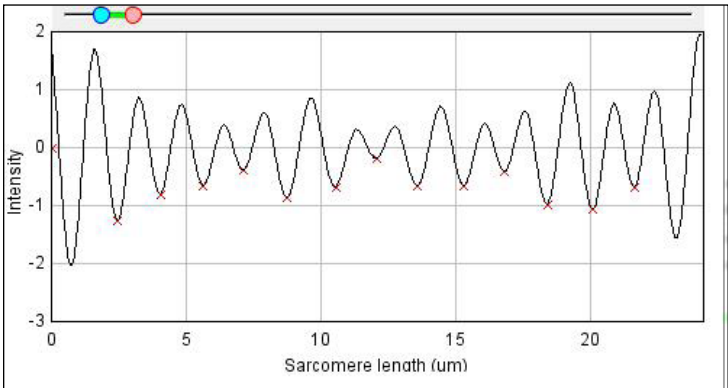


**Fig. 29** Select the DFT filter type to activate the range sliders on the main plugin window.

1. Press  on the main plugin window to begin capturing data.
2. Select the filter frequencies by simply moving the range sliders (Fig. 30). The blue slider (left side) can be used to filter low frequencies, and the red slider (right side) can be used to filter high frequencies. Move the slider to optimize the sinusoidal pattern in the Intensity/Sarcomere Length chart.

NOTE: The slider values are visible in the main Sarcomere Detection window. See Fig. 22.

**TIP:** Move the blue slider (left) to filter out the DC component of the image. Then, move the red slider (right) so that it is close to the blue one. Move the sliders as needed to obtain the best sinusoidal pattern. This filter works best when the sinusoidal pattern is most apparent. The pattern will also depend on your region of interest. Selecting a larger region of interest may require more patience in adjusting this filter.



**Fig. 30** Sinusoidal pattern with the range slider to select the frequencies to be filtered (blue point low frequencies are filtered out; red point high frequencies are filtered out).

**TIP:** Position the blue and red sliders close together for the most stable results ( $\pm 4\text{nm}$ ) when measuring sarcomeres in the  $2\mu\text{m}$  range.



TROUBLESHOOTING

Issue	Possible Cause	Solution
Program will not zoom out.	Only the ROI from the previous is visible, because it wasn't closed properly.	Open the SI-SARCAM-CTS software. There should be an icon on your desktop. Choose the Options menu and select Settings. When the Settings window opens. Press the Reset button.
Plugin crashed before I even started	No ROI was selected before starting the plugin.	Before making any measurements, you must load an image, a video or the live camera field of view and select a region of interest. Then you can start the Sarcomere Detection software.
The time displays with negative values	Occasionally, you may see negative numbers or spikes on the sarcomere length graph.	Close and restart $\mu$ Manager.

**NOTE:** If you have a problem/issue that falls outside the definitions of this troubleshooting section, contact the WPI Technical Support team at 941.371.1003 or [technicalsupport@wpiinc.com](mailto:technicalsupport@wpiinc.com).

SPECIFICATIONS

This unit conforms to the following specifications:

Camera 5MP, monochromatic, USB3, up to 400fps

$\mu$ Manager\* developed at Vale laboratory at UCSF, funded by an NIH grant R01-EB007187 from the National Institute of Biomedical Imaging and Bioengineering (NBIB)

\*Arthur Edelstein, Nenad Amodaj, Karl Hoover, Ron Vale, and Nico Stuurman (2010), Computer Control of Microscopes Using  $\mu$ Manager. Current Protocols in Molecular Biology 14.20.1-14.20.17



## WARRANTY

WPI (World Precision Instruments, Inc.) warrants to the original purchaser that this equipment, including its components and parts, shall be free from defects in material and workmanship for a period of 30 days\* from the date of receipt. WPI's obligation under this warranty shall be limited to repair or replacement, at WPI's option, of the equipment or defective components or parts upon receipt thereof f.o.b. WPI, Sarasota, Florida U.S.A. Return of a repaired instrument shall be f.o.b. Sarasota.

The above warranty is contingent upon normal usage and does not cover products which have been modified without WPI's approval or which have been subjected to unusual physical or electrical stress or on which the original identification marks have been removed or altered. The above warranty will not apply if adjustment, repair or parts replacement is required because of accident, neglect, misuse, failure of electric power, air conditioning, humidity control, or causes other than normal and ordinary usage.

To the extent that any of its equipment is furnished by a manufacturer other than WPI, the foregoing warranty shall be applicable only to the extent of the warranty furnished by such other manufacturer. This warranty will not apply to appearance terms, such as knobs, handles, dials or the like.

WPI makes no warranty of any kind, express or implied or statutory, including without limitation any warranties of merchantability and/or fitness for a particular purpose. WPI shall not be liable for any damages, whether direct, indirect, special or consequential arising from a failure of this product to operate in the manner desired by the user. WPI shall not be liable for any damage to data or property that may be caused directly or indirectly by use of this product.

## Claims and Returns

Inspect all shipments upon receipt. Missing cartons or obvious damage to cartons should be noted on the delivery receipt before signing. Concealed loss or damage should be reported at once to the carrier and an inspection requested. All claims for shortage or damage must be made within ten (10) days after receipt of shipment. Claims for lost shipments must be made within thirty (30) days of receipt of invoice or other notification of shipment. Please save damaged or pilfered cartons until claim is settled. In some instances, photographic documentation may be required. Some items are time-sensitive; WPI assumes no extended warranty or any liability for use beyond the date specified on the container

Do not return any goods to us without obtaining prior approval and instructions from our Returns Department. Goods returned (unauthorized) by collect freight may be refused. Goods accepted for restocking will be exchanged or credited to your WPI account. Goods returned which were ordered by customers in error are subject to a 25% restocking charge. Equipment which was built as a special order cannot be returned.

## Repairs

Contact our Customer Service Department for assistance in the repair of apparatus. Do not return goods until instructions have been received. Returned items must be securely packed to prevent further damage in transit. The Customer is responsible for paying shipping expenses, including adequate insurance on all items returned for repairs. Identification of the item(s) by model number, name, as well as complete description of the difficulties experienced should be written on the repair purchase order and on a tag attached to the item.

*\* Electrodes, batteries and other consumable parts are warranted for 30 days only from the date on which the customer receives these items.*

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